

## Whitefly, Host Plant and Parasitoid: A Review on Their Interactions

### M Mostafizur Rahman Shah<sup>1</sup>, Shi-Ze Zhang<sup>2</sup>, Tong-Xian Liu<sup>3</sup>

<sup>1</sup>Senior Scientific Officer, Division of Entomology, Bangladesh Agricultural Research Institute, Rangpur, Bangladesh
<sup>2</sup>Asst. Professor, College of Plant Protection, Northwest A&F University, Shaanxi, China
<sup>3</sup>Professor & Dean, College of Plant Protection, Northwest A&F University, Shaanxi, China

ARTICLE INFO	ABSTRACT
Volume 4 Number 1/2015 Issue 10 DOI Prefix: 10.15590 Received: Feb 17, 2015 Accepted: Feb 20, 2015 Published: Feb 22, 2015 E-mail for correspondence: mostafiz.wrc@gmail.com	The whitefly, <i>Bemisia tabaci</i> (Gennadius) is an enormously polyphagous insect pest showing the intercrop movement, high reproduction, resistances to insecticides, under-leaf habitat, and virus transmission. In the past decades, numerous studies revealed the interactions of whitefly, host plant, and their natural enemies. The oviposition, development, survivorship, and behavior of <i>B. tabaci</i> are mostly mediated by host plants. Numerous parasitoid species are essential components controlling whiteflies biologically; they parasitize whitefly nymphs and kill them through active feeding. Studies showed that appropriate conservation and augmentation techniques of parasitoids can make them more dynamic on the crops in protected environment and even in exposed cropping field. Moreover, the performance of parasitoid is influenced by several factors including number and release rate of parasitoid, intra- and inter-specific competition, stage, size and density of whitefly nymph as well as environmental factors and host plants. In this review, we summarized the progress and findings of how <i>B. tabaci</i> adapt on various plant species, interactions among host plant, <i>B. tabaci</i> and parasitoid in the last decade. We also discussed the possibility of using parasitoids for suppression of whitefly populations.

Source of Support: Nil, Conflict of Interest: None Declared.

How to Cite: Shah MM, Zhang S and Liu T. 2015. Whitefly, Host Plant and Parasitoid: A Review on Their Interactions Asian Journal of Applied Science and Engineering, 4, 48-61.

This article is is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License. Attribution-NonCommercial (CC BY-NC) license lets others remix, tweak, and build upon work non-commercially, and although the new works must also acknowledge & be non-commercial.



#### INTRODUCTION

The whitefly, *Bemisia tabaci* (Gennadius) is one of the most important insect pests of most vegetables, some ornaments and agronomic crops especially in the sub-tropical and tropical regions around the world (Liu, 2007; Wan et al., 2009; Zhang et al., 2007). The whitefly has association with almost 600 different species of plants (Martin et al., 2000; Naranjo and Ellsworth, 2009; Secker et al., 1998) which comprise large number of cultivated and non-cultivated, annuals and perennials (Bedford et al., 1992; Brown et al., 1995; Butler and Henneberry, 1986). Systematic studies of the *B. tabaci* reveal convincing evidence that *B. tabaci* is a complex species including at least 35 cryptic species with extensive genetic diversity (Ahmed et al., 2013; Bing et al., 2013; De Barro et al., 2011; Dinsdale et al., 2010; Hu et al., 2011; Liu et al., 2012; Wang et al., 2011; Xu et al., 2011) which show diverse behavior concerning to host plant preference, oviposition, ecological adaptation as well as virus dissemination (De Barro et al., 2005; Perring, 2001). Furthermore, it can be simply adapt to the new host plants in the new area because of its distinctive biological and behavioral characteristics.

With the introduction of synthetic chemical insecticides in the 1950s, pest control has been mostly relied on that; however, it soon became noticeable that there were problems associated with the use of insecticides. At present, the management of insect and mite pests in agricultural systems still relies primarily on the use of chemicals, and consequently, many insect pests became resistant to chemical insecticides, including the whitefly *B. tabaci* (Brun-Barale et al., 2010; Byrne et al., 2010; Mallet, 1989; Schuster et al., 2010). In addition, the extensive use of insecticides adversely affected non-target organisms and caused secondary pest resurgence with environmental and health concerns.

Establish a long lasting integrated pest management system using efficient natural enemies is an encouraging decision. Parasitoids have drawn significant attention because of their importance in biological control of insect pests through parasitism and host feeding (Chan and Godfray, 1993). The host feeding mostly involves in consumption of host fluids, secreted during oviposition which has reported in at least 140 species belonging 17 families of Hymenoptera (Jervis and Kidd, 1986; Zang and Liu, 2010; Zang et al., 2011a,b). About 10,000 wasp species feed their host (Kidd and Jervis, 1989) that supplies nutrients to the parasitoids assisting egg development (Heimpel and Collier, 1996; Jervis and Kidd, 1986) and consequently increases parasitoid's egg production (Burger et al., 2005; Giron et al., 2004; Rivero and Casas, 1999; Ueno, 1999). In view of that consideration, host feeding activities believed an encouraging attribute of parasitoids as selecting a candidate for biological control agent (Jervis and Kidd, 1986; Zang and Liu, 2010; Zang et al., 2011a). Whiteflies show multivoltine characters, they have no diapauses or dormant stages. Therefore, whiteflies are constant through the frequent utilization of numerous plant host resources over the annual cycle (Naranjo et al., 2009). Movement from one plant host species to another is consequently an essential element of the whitefly ecology which facilitates finding host plants and colonization in a frequently variable ecosystem. In this paper, we reviewed the recent evolution and findings of adaptation of B. tabaci on various plants and ecosystem, and their interactions with parasitoids in different aspects in the last decade. We hope that this review will provide comprehensive information about B. tabaci, host plant, and parasitic natural enemies and to stimulate more research in these needed areas.

#### ADAPTATION OF BEMISIA TABACI TO VARIOUS HOST PLANTS AND CROP SYSTEM

While and wherever a female insect lays eggs or deposits larvae have a determining influence on her fitness as well as adaptation. Feeding and oviposition are crucial for host selection in herbivores that enhance them to adapt to a new host. It was thought that both the plant nutrients and secondary volatile chemicals might be the most critical for insects to establish them on a particular host. However, whiteflies have ability to adapt to new host plants and zones for the reasons of its sole biochemical and biological characteristics. Carabali et al. (2005) believed that B. tabaci acclimate potentially in the Neotropics that consider a great threat for cassava cultivation. To confirm their assumption, Carabali et al. (2005) carried out an experiment whether B. tabaci B biotype can establish on the cassava plant (Manihot esculenta L.). They found that B. tabaci was becoming establish on M. esculenta, sequentially started from Phaseolus vulgaris L. followed by Euphorbia pulcherrima Willd. and Jatropha gossypiifolia L. In that sequential adaptation, the highest survival rate, and the lowest developmental duration were needed for those whiteflies coming from J. gossypiifolia as closest phylogenetic host plant. Conversely, Hu et al. (2011) explained that poinsettia derived Q biotype B. tabaci is unable to establish rapidly on field crops though it might have some potentiality to adapt.

Population dynamics of B. tabaci are regulated primarily by predation (Naranjo and Ellsworth, 2005) as well as by abiotic factors (mostly by temperature) in the field (Naranjo et al., 2003). To concentrate on the population dynamics, Liu (2000) observed few B. tabaci populations on collards in winter and did not show any economic significance damage at relatively low temperature as compared with high level of infestation found in the fall and spring. Lin et al. (2007) investigated the overwintering of B. tabaci with its population dynamics in northern China, a cold region that how they survive a winter. To address that research, they conducted a cold hardiness test and found that in northern China it is impossible to survive for natural B. tabaci populations under natural environment during winter season. Furthermore, they observed that population density of B. tabaci was lower at winter and gradually became higher during February-March until they established in the field in the month of April. On the other hand, high temperature is also a reasonable cause for low population densities of ectothermic organisms. Survival of B. tabaci adults and hatching of  $F_1$  nymphs were critically affected when whiteflies were exposed to 41°C or higher (Cui et al., 2008), although this study revealed that B. tabaci have capability to tolerate at higher temperature than that of Trialeurodes vaporariorum (Westwood).

#### INTERACTIONS BETWEEN BEMISIA TABACI AND HOST PLANTS

*B. tabaci* infests at least 600 different plant species, though however, it showed preference to Asteraceae, Compositae, Cruciferae, Cucurbitaceae, Euphorbiaceae, Fabaceae, Labiatae, Leguminosae, Malvaceae and Solanaceae families' plants (Bayhan et al., 2006; Elsey and Farnham, 1994; Johnson et al., 1982; Mound and Halsey, 1978), with a distinct affinity to cabbage, cotton, cucumber, eggplant, gerbera, gherkin, melon, poinsettia, squash, sweet potato, and tomato (van Lenteren and Noldus, 1990; Shah and Liu, 2013; Xu et al., 2011; Ying et al., 2003). *B. tabaci* exhibited oviposition choice among different host plants as well as leaf surfaces within a host (Gruenhagen and Perring, 2001a; Liu and Stansly, 1995).

Semiochemicals released by host plants known as kairomones, play a significant role in mediating host choice on the majority of the herbivorous insects (Xiao and Fadamiro, 2009). Lee et al. (2011) described that whitefly adult significantly preferred cucumber to poinsettia, when whiteflies were given to choose between those two host plants.

Asian Business Consortium | AJASE • Feb 2015 • Vol 4 • Issue 10

Nevertheless, if the whiteflies had first established on poinsettia, adults were not attracted successfully on cucumber from poinsettia plants. Even adequate number of adults was not attracted on eggplant, though adults showed preference to eggplant in the primary assay and where offered a choice between eggplant and poinsettia (Lee et al., 2009). This phenomenon reflects the behavioral attributes of adult whiteflies. Similar result was stated by Bird and Krüger (2006) where female B. tabaci adult stayed on the same host, and very few movements among host plants were recorded during their observation. Omondi et al. (2005) examined the host preference between two biotypes of B. tabaci namely cassava and okra biotype, where cassava biotype significantly chose cassava for oviposition and habitation rather than okra, Abelmoschus esculentus L. In contrast, okra populations have had preference on okra and also laid eggs on eggplant, Solanum melongena L., tomato, Solanum lycopersicum L., garden eggplant, S. integrifolium Poir and that on cowpea, Vigna unguiculata L., but not on cassava plant. The offspring of okra population developed suitably in almost all host species except for that on cassava and slightly subsisted on pepper, Capsicum annum L and cabbage, Brassica oleracea L. Shah and Liu (2013) established whitefly populations on eggplant, tomato and cucumber where they found that host preferences were influenced by both whitefly populations and host plants however host plants played stronger role.

The whitefly, *B. tabaci* perform differently on different host species, even though sometimes differs when feeding on dissimilar varieties within a same plant species. Among several varieties or lines of melon (*Cucumis melo* L.), Liu (2003) observed in choice test that fewer *B. tabaci* adults along with eggs were observed on "Hymark" than that on "TAM Sun", "TAM Sun × gl" and "Primo". On the other hand, more eggs were found on "Hymark" and "Primo" than those on "TAM Sun" and "TAM Sun × gl" in no-choice test, but whitefly immature mortality was considerably higher on "Hymark". "Primo" had the utmost numbers of adults and pupae, and also immature developed significantly faster on it than on other varieties. Baldin and Beneduzzi (2010) investigated the host preference of *B. tabaci* biotype B among squash (*Cucurbita pepo* L.) varieties/ lines where the least number of adults was attracted on "Bianca" in choice test. In choice tests, the adult did not prefer to oviposit on the varieties "Novita Plus", "Bianca" and "Sandy" in no-choice test. However, "Sandy" expressed the highest mortality against whitefly nymphs.

Whitefly feeding can induce a defensive response on host plants to the other competing herbivores. Several factors are responsible for the quantity and quality of defensive induction including host plant species and varieties, their growing environments, duration of whitefly feeding on host plants, whitefly population density, etc. Mayer et al. (2002) observed behavioral differences between two herbivores, *Trichoplusia ni* (Hübner) and *Liriomyza trifolii* (Burgess). When these herbivores exposed on plants that were previously infested by *B. tabaci* have shown inferior feeding and oviposition, reduced development and survival. Whereas, a little or no affect on whitefly's feeding and oviposition behaviors when the whitefly was introduced to the plants previously exposed to the leaf miners.

#### INTERACTIONS BETWEEN BEMISIA TABACI AND PARASITOIDS

Parasitoids are primarily important agents to manage whiteflies biologically; they parasitize whitefly nymphs to produce their new generations and also feed the whitefly nymphs to improve their fitness. *Encarsia* and *Eretmocerus* are the fundamental genera among the widespread fauna of the *B. tabaci* parasitoids. Gerling et al. (2001) reported 34 *Encarsia* spp., 12 *Eretmocerus* spp., two *Amitus* spp., and one from each *Metaphycus* spp. and *Signiphora* spp.

of *B. tabaci* parasitoids. Besides the Gerling et al. (2001) list, Arnó et al. (2010) further reported 15 *Encarsia* spp. and 10 *Eretmocerus* spp. Childs et al. (2011) also reported eight species of both *Encarsia* and *Eretmocerus* genera from Western Sydney, among them one species was not included either in the list of Gerling et al. (2001) or in that of Arnó et al. (2010). Li et al. (2011) found 41 *Encarsia* spp., 12 *Eretmocerus* spp., two *Amitus* spp. and one *Ablerus* sp. in their survey in south, southeast, north, northwest, and central China.

The majority researches have revealed that premature hosts are usually preferred by parasitoids for feeding, whereas some researches revealed that the mature nymphs are preferred, though host size did not affect on the host feeding (Antolin, 1989; Heimpel and Rosenheim, 1995; Mansaray and Sundufu, 2010; Videllet et al., 1997). It has been examined that Er. melanoscutus Zolnerowich and Rose and En. pergandiella Howard females parasitized the most on the third instar nymphs and the lowest on the first instar nymphs of B. tabaci (Greenberg et al., 2008; Jones and Greenberg, 1999; Liu, 2007) whereas Er. mundus Mercet was most successfully parasitized on younger host instars (Ghahari et al., 2005; Greenberg et al., 2008). Zang and Liu (2008) showed that the host feeding capacities were significantly different among En. formosa Gahan, En. sophia (Girault & Dodd) and Er. melanoscutus when offered them either on single instar or diverse instars where En. sophia showed better performance for feeding host than that of others. Xiao et al. (2011) reported that En. sophia has strong parasitization capacity on both Trialeurodes variabilis (Quaintance) and B. tabaci. Yang and Wan (2011) reported that Er. hayati Zolnerowich & Rose effectively parasitized and fed almost all instar nymphs of *B. tabaci* except the late fourth instar; parasitism on first, second, and third instars were significantly higher than on fourth instar but the utmost prevalence of parasitism was shown on the first instar. Younger nymphal instars were fed on by female parasitoids than older ones. The highest development time was needed for Er. hayati when parasitized on the first instar, and intermediate on the second and third instars but lowest on early fourth instar.

Adult parasitoids gain nutrients from the host through feeding insight it, but this process destroys oviposition opportunity on host. Honeydew considered a nutrient rich substitute for the Hemipteran parasitoids. Studies revealed that host secreted honeydew as substitute nutrition increased egg quantity and adult longevity of parasitoids (Burger et al., 2004); whereas host feeding might facilitate female parasitoids to continue the highquality hydropic egg production that is essential for natural environment. Burger et al. (2005) observed that parasitoids can be acquired advantage and increased their oviposition rate through host feeding compared to honeydew feeding. This task might be relevant under circumstances of high host population density or high mortality of parasitoid adults and if other sources of food may not provide sufficient nutrition for egg production. Almost all parasitoids kill whitefly nymphs either by piercing the nymph with ovipositor following egg oviposition or by piercing the nymph and sucking the body fluids through labium (Shi et al., 2009; Zang and Liu, 2009). The efficacy of the parasitoid performance for controlling whiteflies might be increased by food deprivation for an optimal period prior to release. Zang and Liu (2009) reported that En. sophia feeds and parasitizes more B. tabaci nymphs during their whole lifespan if the adult parasitoids keep on without food for 6 hours prior to release and found that they live considerably longer than the parasitoids of no food deprivation. Zang and Liu (2010) also examined on Er. melanoscutus and En. formosa and found that parasitoids depriving food for 6 hours feeds more B. tabaci nymphs and parasitizes equal or more number than the parasitoids of continuous feeding.

Studies revealed that parasitoids show preference to oviposit on larger hosts for their immature development suitably though smaller host is better to control the host insect effectively. Luo and Liu (2011) described that the adults of *En. sophia* were larger those were emerged from *T. vaporariorum* rather than from *B. tabaci*. Where all parameters of biological assays except the development time were superior when parasitized larger host than smaller host. Besides, the adult wasps parasitized more nymphs emerging from *T. vaporariorum* than those from *B. tabaci*.

Zang et al. (2011a) have found in their experiment that when they offered *Er. melanoscutus* and *En. sophia* in different densities of *B. tabaci* nymphs the sexed females parasitized and host-fed more number of whitefly nymphs at any host density than unsexed females. Both the females of *Er. melanoscutus* either sexed or unsexed parasitized and host-fed more whitefly nymphs in high host density population. However, sexed females of *En. sophia* fed more whitefly nymphs at high density of host population, but no nymphs were parasitized by unsexed females.

Parasitoids have direct effects selecting the 'right' hosts on the success of reproduction and effectiveness as an agent to control whiteflies. Gelman et al. (2005) stated the physiological and biochemical relations between parasitoids and whiteflies, and they assumed that the parasitoids inject and/or generate bio-chemicals inside their hosts that interfere with the immune system of the host. They also observed that parasitoids maneuver host maturity according to their own needs. Besides, parasitoids must synchronize their own development with that of their hosts, even though if their eggs were laid in fourth instar nymphs.

#### INTERACTIONS BETWEEN PARASITOIDS AND PLANTS

Whiteflies are polyphagous in nature, and its host plants may mediate the activity of natural enemies noticeably (Inbar and Gerling, 2008; Mansaray and Sundufu, 2010). Plant species and varieties comprise plant attributes including leaf features, volatile compounds, presence or absence of pubescence and/or wax, etc. High dense and rigid hairs on the leaves generate low rates of parasitism (Rajam et al., 1988). Pubescent and trichomes on the leaves reduce parasitism of whiteflies (Gruenhagen and Perring, 2001b; Oster, 1995). *Eretmocerus* species associate greater parasitism rate on hirsute varieties of melon and soybean (Headrick et al., 1996a; McAuslane et al., 2000), indicating that some parasitoid species might be more efficient on certain pubescent plant leaves. Parasitism of the whitefly, *B. tabaci* was poorer on velvetleaf, *Aboutilon theofrasti* Medic than that of on other plants due to discharge of trichome exudates that captured parasitoids (Gruenhagen and Perring, 2001b). Waxy leaf surface also affects parasitism and *Eretmocerus* spp. and *En. pergandiella* had a greater parasitism on reduced wax both in laboratory and field crops than on other plants (Jackson et al., 2000; McAuslane et al., 2000).

#### **INTERACTIONS AMONG PARASITOIDS**

Interspecific or intraspecific competitive interactions act on the determining of the function and structure of ecological communities. A plenty of parasitoid species are available in the field. Multiple species of parasitoids attack single host species. Competition among parasitoids could significantly decrease their control efficacy due to inhibition of their reproduction causing from competitive multi-parasitization or heterospecific ovicides. Existence of diversity of parasitoid species might influence negative impact on pest management system if they coexist in the same system for a long period. Stansly et al. (2004, 2005a) reported that *Er. mundus* has higher parasitism efficiency and more ability to manage B. tabaci than Er. eremicus Rose & Zolnerowich in coexisting ecosystem of parasitoids. They observed that Er. mundus quickly replaced Er. eremicus from greenhouses when both of them released together or solely *Er. eremicus*, suggesting that a huge number of Er. mundus entered into the greenhouses from the outer area. Collier and Hunter (2001) reported lethal effect of Er. eremicus with En. sophia reflecting a combination of multiparasitism and host feeding on parasitized hosts. It is predicted that interruption might happen while Er. eremicus direct interacted on En. sophia through a lethal interference interaction, for example, with intrinsic competition (multi-parasitism) or facultative hyperparasitism. These results reveal that competitive interactions among parasitoids may cause displacement of inferior species. Therefore, using of multiple parasitoid species at a time might not be successful to control whitefly (Collier et al., 2002). In contrast, other researchers suggested that competitive interactions among parasitoid species do not interrupt the suppression of host population and, even though, facilitate biological control. It is revealed that though the populations of primary parasitoid were reduced by the interference of auto-parasitoid but had no concomitant disturbance on host suppression suggesting that an auto-parasitoid might be much effective for suppressing host populations and might enhance the whitefly management program (Bográn et al., 2002; Heinz and Nelson, 1996; Hunter et al., 2002; Zang et al., 2011b). In the competitive interactions among parasitoids, one species can affect the other species directly by means of multi-parasitism whereas intra-specific interference or inter-specific interference through host-feeding of the parasitized hosts (Collier and Hunter, 2001; Zang et al., 2011b), while heterospecific ovicides may happen (Collier et al., 2007; Pedata et al., 2002). However, Pang et al. (2011) studied the interspecific competitive interactions of En. formosa and En. sophia where found that the parasitization rate by En. formosa was higher than that of En. sophia while used both of them individually. In contrast, the number of parasitized nymphs of B. tabaci was reduced considerably when both the Encarsia species was applied together either simultaneous or sequential flow compared to individual application. However, dual application of the parasitoids killing more *B. tabaci* nymphs than that of a single application. Therefore, though the offspring of the parasite will be reduced by interspecific competition, but this competition may not affect the efficiency of whitefly control.

Super-parasitism and multi-parasitism delay the developmental time of parasitoids' offspring and decrease larval survivorship, especially in solitary parasitoids. So, discrimination behavior during host selection, i.e. differentiating un-parasitized nymphs instead of parasitized nymphs is important to avoid killing time, parasitization and to minimize the mortality rate of their progeny. Thus, host discrimination behavior in intraspecies interaction is frequently observed in parasitoids having solitary in nature to prevent competition among their own offspring, whereas that behavior is just only a few in interspecific interactions (Agboka et al., 2002; Royer et al., 1999). Ardeh et al. (2005) examined the host preference of Er. eremicus and Er. mundus, and found that superparasitism were escaped by experienced female. Conversely, naïve females laid eggs on the parasitized hosts, which already parasitized by conspecific females. The hosts parasitized previously by Er. mundus females, were not multi-parasitized by Er. eremicus. Conversely, Er. mundus multi-parasitized the nymphs that had already been parasitized by Er. eremicus. Neither Er. mundus nor Er. eremicus was stronger in case of super-parasitism, even though Er. mundus was stronger than Er. eremicus in case of multi-parasitism. Huang et al. (2009) examined hyperparasitism of naïve female En. tricolor (Förster) through direct behavioral observation using En. formosa, a heterospecific parasitoid, and En. tricolor, a conspecific parasitoid as the secondary host at their different larval stages. Encountering rate of recurrence by *En. tricolor* at all larval stages was similar on both host species. Nonetheless, hyperparasitism varied on different larval stages of different hosts, where late larval stages and heterospecific parasitoid, *En. formosa* was most preferred. *En. tricolor* behavior concerned searching, and hyperparasitism was independent of preference to the different nymphal stages of whitefly (*Aleyrodes proletella* L.) (Huang et al., 2009). Similar study was conducted by Zang et at. (2011b), where they stated that *En. Sophia*, an autoparasitoid, preferred heterospecifics as secondary hosts to produce males. Their study also stated that the females of *En. sophia* parasitized more hosts if they mated with the heterospecifics derived males than the females mated with the conspecifics derived males.

#### INTERACTIONS AMONG BEMISIA TABACI, HOST PLANTS AND PARASITOIDS

The traits of host plant may alter interactions between herbivores and parasitoids by influencing the herbivores and parasitoids while infesting on host plants. De Barro et al. (2000) verified that *Er. mundus* originating from cotton (*Gossypium hirsutum* L.) parasitized more *Bemisia tabaci* nymphs or also parasitized more if tested on rockmelon (*Cucumis melo* L.), while lower parasitism was observed in both cases either the parasitoids were originating from tomato or tested on tomato. They assumed that the rough surface of leaves enhance parasitism due to preventing fitting level of nymph underside on the leaf surface which make easier to parasitoids placing their eggs underside the hosts that previously examined by Headrick et al. (1996b). De Barro et al. (2000) also supposed, long, and dense hairs on tomato leaf surface interfere parasitoids movement resulting reduce parasitism that as well stated by van Lenteren et al. (1995).

Herbivores diet of different qualities can mediate differently on the interactions involving plants-herbivores-parasitoids (Harvey et al., 2003). Greenberg et al. (2002) stated that Er. mundus developed faster and produced more offspring, and its parasitism and emergence rate was greater on B. tabaci than that on T. vaporariorum onto tomato whereas, the performance of *Er. eremicus* was similar in both the species except for its female deposited more eggs in B. tabaci nymph than that of T. vaporariorum nymph. They also reported that T. vaporariorum emerged female offspring of both Er. mundus and Er. eremicus were considerably larger than those of B. tabaci. Urbaneja et al. (2007) reported the adult longevity of Er. mundus was shorter in the second instar of B. tabaci on tomato than those of sweet pepper where the fecundity was more on sweet pepper than that on tomato. They also stated that host feeding incidence of Er. mundus was noticeably higher on sweet pepper than that of tomato, and the net reproductive rate ( $R_0$ ) was greater on tomato (63.8) than that of sweet pepper (51.0). Greenberg et al. (2009) described cotton was the most favorable to sweetpotato whitefly, and bean was the most favorable to greenhouse whitefly. They observed that preimaginal mortality of sweetpotato whitefly on cotton was less and developmental time also shorter than that of greenhouse whitefly. Parasitism by Er. mundus was greatest on sweetpotato whitefly and least on greenhouse whitefly when both whiteflies were reared on cotton, whereas parasitism by En. pergandiella was greatest on sweetpotato whitefly reared on bean or cotton. Emergence of Er. mundus was the greatest from sweetpotato whitefly on cotton, where emergence of En. pergandiella was noticeably greater than that of *Er. mundus* among host plants.

Release rate of parasitoids mostly depends on plant host species, growth stage of plants, parasitoid species, herbivore density, field condition and area of crop field. At low density of herbivore, parasitoids release periodically at lower rate showed better performance than

once release at higher rate (Gu et al., 2008). Stansly et al. (2005b) observed on pepper plants that whitefly populations, both nymphs and adults were reduced over 90% when released parasitoids at both high and low rates (6 *Er. mundus*/m<sup>2</sup>/week, 1.5 *Er. mundus*/m<sup>2</sup>/week, respectively). Whitefly densities of both nymphs and adults were decreased exceeding 95% on tomato plants due to higher rate of parasitoid initiation but about 54% nymphs and 69% adults at the lower rate. Similar results were described by Bellows et al. (2006), whereas Qiu et al. (2004) reported 3-5 females of *Eretmocerus* sp. per cucumber plant is optimal for controlling of *B. tabaci*. However, Lopez and Andornoa (2009) observed that at the rate of two *Er. mundus*/plant/week was sufficient to manage whitefly population on pepper but if parasitoids had to be released three times achieving the best pest suppression with 84% of parasitism.

#### CONCLUSION

The whitefly, *B. tabaci* establishment onto new host plant is mainly considered by feeding as well as oviposition preference which is mediated by plant leaf features and chemicals inside. Moreover, natural enemies with abiotic factors play a fundamental role to adapt whitefly on host plants. Parasitoids prefer larger host to oviposit and show better performance but more efficient on smaller host to control. A large number of researches revealed, parasitoids usually prefer earlier host stages to feed, and only some studies illustrated that older host stages are preferred. Some parasitoids have higher parasitism ability, and their efficiency depends on augmentation period. Therefore, during application of parasitoid species, sequence of parasitoid initiation, intra-specific or inter-specific competitive interactions of parasitoids, host plant species, whitefly population density and nymphal stages, and finally field condition. To develop the best strategies for whitefly management on vegetables and other field crops, the careful thought should be given to the interactions among three components including whitefly-host plant-parasitoid.

#### REFERENCES

- Agboka, K., Schulthess, F., Chabi-Olaye, A., Labo, I., Gounou, S., Smith, H., 2002. "Self-, intra-, and interspecific host discrimination in *Telenomus busseolae* Gahan and *T. isis* Polaszek (Hymenoptera: Scelionidae), sympatric egg parasitoids of the African cereal stem borer *Sesamia calamistis* Hampson (Lepidoptera: Noctuidae)". J. Insect. Behav. 15: 1-12.
- Ahmed, M.Z., De Barrow, P.J., Ren, S.X., Greeff, J.M., Qiu, B.L., 2013. Evidence for horizontal transmission of secondary endosymbionts in the *Bemisia tabaci* cryptic species complex. *PLoS ONE* 8, e53084.
- Antolin, M.F., 1989. Genetic considerations in the study of attack behavior of parasitoids, with reference to *Muscidifurax raptor* (Hymenoptera: Pteromalidae). *Fla. Entomol.* 72: 15-32.
- Ardeh, M.J., de Jong, P.W., van Lenteren, J.C., 2005. Intra- and interspecific host discrimination in arrhenotkous and thelytokous *Eretmocerus* spp. *Biol. Control* 33: 74-80.
- Arnó, J., Gabarra, R., Liu, T.X., Simmons, A.M., Gerling, D., 2010. Natural Enemies of Bemisia tabaci: Predators and Parasitoids, in: Stansly, P.A., Naranjo, S.E. (Eds.), Bemisia: Bionomics and Management of a Global Pest. New York, Springer, London, pp. 385-422.
- Baldin, E.L.L., Beneduzzi, R. A., 2010. Characterization of antibiosis and antixenosis to the whitefly silverleaf *Bemisia tabaci* B biotype (Hemiptera: Aleyrodidae) in several squash varieties. J. Pest Sci. 83: 223-229.
- Bayhan, E., Ulusoy, M.R., Brown, J.K., 2006. Host range, distribution, and natural enemies of *Bemisia tabaci* 'B biotype' (Hemiptera: Aleyrodidae) in Turkey. J. Pest Sci. 79: 233-240.
- Bedford, I.D., Briddon, R.W., Markham, P.G., Brown, J.K., Rosell, R.C., 1992. *Bemisia tabaci*-biotype characterization and the threat of this whitefly species to agriculture, in: Brighton Crop Prot. Conf.-Pests and Diseases. British Crop Protection Council, Farnham, UK, pp. 1235-1240.

Asian Business Consortium | AJASE • Feb 2015 • Vol 4 • Issue 10

- Bellows Jr T.S., Paine, T.D., Bezark, L.G., Ball, J., 2006. Optimizing natural enemy release rates, for *Encarsia inaron* Walker (Hymenoptera: Aphelinidae) and *Siphoninus phillyreae* (Haliday) (Homoptera: Aleyrodidae). *Biol. Control* 37: 25-31.
- Bing, X.L., Yang, J., Zchori-Fein, E., Wang, X.W., Liu, S.S., 2013. *Hemipteriphilus asiaticus*, a newly-described whitefly symbiont, in: First International Whitefly Symposium, Kolymbari, Crete, Greece. p. 83.
- Bird, T.L., Krüger, K., 2006. Response of the polyphagous whitefly *Bemisia tabaci* B-biotype (Hemiptera: Aleyrodidae) to crop diversification influence of multiple sensory stimuli on activity and fecundity. *Bull. Entomol. Res.* 96: 15-23.
- Bográn, C.E., Heinz, K.M., Ciomperlik, M.A., 2002. Interspecific competition among insect parasitoids: field experiments with whiteflies as hosts in cotton. *Ecology* 83: 653-668.
- Brown, J.K., Frohlich, D.R., Rosell, R.C., 1995. The sweetpotato or silverleaf whiteflies: biotypes of Bemisia tabaci or a species complex? Annu. Rev. Entomol. 40: 511-534.
- Brun-Barale A., Héma, O., Martin, T., Suraporn, S., Audant, P. et al., 2010. Multiple P450 genes overexpressed in deltamethrin-resistant strains of *Helicoverpa armigera*. *Pest Manag. Sci.* 66: 900-909.
- Burger, J.M.S., Kormany, A., van Lenteren, J.C., Vet, L.E.M., 2005. Importance of host feeding for parasitoids that attack honeydew-producing hosts. *Entomol. Exp. Appl.* 117: 147-154.
- Burger, J.M.S., Reijnen, T.M., van Lenteren J.C., Vet, L.E.M., 2004. Host feeding in insect parasitoids: why destructively feed upon a host that excretes an alternative? *Entomol. Exp. Appl.* 112: 207-215.
- Butler, G.D., Henneberry, T.J., 1986. *Bemisia tabaci* (Gennadius), a pest of cotton in the southwestern United States. *Agric. Res. Serv. Tech. Bull.* 19: 1701.
- Byrne, F.J., Oletting, R.D., Bethke, J.A., Green, C., Chamberlin, J. 2010. Understanding the dynamics of neonicotinoid activity in the management of *Bernisia tabaci* whiteflies on poinsettias. *Crop Prot.* 29: 260-266.
- Carabali, A., Bellotti, A.C., Montoya-Lerma, J., Cuellar, M.E., 2005. Adaptation of *Bemisia tabaci* biotype B (Gennadius) to cassava, *Manihot esculenta* (Crantz). *Crop Prot.* 24: 643-649.
- Chan, M.S., Godfray, H.C.J., 1993. Host-feeding strategies of parasitoid wasps. Evol. Ecol. 7: 593-604.
- Childs, I.J., Gunning, R.V., Mansfield, S., 2011. Parasitoids of economically important whiteflies associated with greenhouse vegetable crops in western Sydney. *Aust. J. Entomol.* 50: 296-299.
- Collier, T.R., Hunter, M.S., 2001. Lethal interference competition in the whitefly parasitoids *Eretmocerus eremicus* and *Encarsia sophia*. Oecologia 129: 147-154.
- Collier, T.R., Hunter, M.S., Kelly, S.E., 2007. Heterospecific ovicide influences the outcome of competition between two endoparasitoids, *Encarsia formosa* and *Encarsia luteola*. *Ecol. Entomol*. 32: 70-75.
- Collier, T.R., Kelly, S.E., Hunter, M.S., 2002. Egg size, intrinsic competition, and lethal interference in the parasitoids *Encarsia pergandiella* and *Encarsia formosa*. *Biol. Control* 3: 254-261.
- Cui, X.H., Wan, F.H., Xie, M., Liu, T.X., 2008. Effects of heat shock on survival and reproduction of two whitefly species, *Trialeurodes vaporariorum* and *Bemisia tabaci* biotype B. J. Insect Sci. 8: 24.
- De Barro, P.J., Hart, P.J., Morton, R., 2000. The biology of two *Eretmocerus* spp. (Haldeman) and three *Encarsia* spp. Forster and their potential as biological control agents of *Bemisia tabaci* biotype B in Australia. *Entomol. Exp. Appl.* 94: 93-102.
- De Barro, P.J., Liu, S.S., Boykin, L.M., Dinsdale, A.B., 2011. *Bemisia tabaci*: A statement of species status. *Annu. Rev. Entomol.* 56: 1-19.
- De Barro, P.J., Trueman, W.H., Frohlich, D.R., 2005. *Bemisia argentifolii* is a race of *B. tabaci* (Hemiptera: Aleyrodidae): the molecular genetic differentiation of *B. tabaci* populations around the world. *Bull. Entomol. Res.* 95: 193-203.
- Dinsdale, A., Cook, L., Riginos, C., Buckley, Y.M., De Barro, P.J., 2010. Refined global analysis of *Bemisia tabaci* (Hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) mitochondrial cytochrome oxidase I to identify species level genetic boundaries. *Ann. Entomol. Soc. Am.* 103: 196-208.
- Elsey, K.D., Farnham, M.W., 1994. Response of *Brassica oleracea* L. to *Bemisia tabaci* (Gennadius). *Hort. Sci.* 29, 814-817.
- Feng, Y.T., Wu, Q.J., Wang, S.L., Chang, X.L., Xie, W. et al., 2010. Cross-resistance study and biochemical mechanisms of thiamethoxam resistance in B biotype *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Pest Manag. Sci.* 66: 313-318.
- Gelman, D.B., Gerling, D., Blackburn, M.B., Hu, J.S., 2005. Host-parasite interactions between whiteflies and their parasitoids. Arch. Insect Bioch. Physiol. 60, 209-222.

- Gerling, D., Alomar, O., Arnó, J., 2001. Biological control of *Bemisia tabaci* using predators and parasitoids. *Crop Prot.* 20: 779-799.
- Ghahari, H., Huang, J., Wang, Z.H., 2005. Biology of a thelyotokous biotype of *Eretmocerus mundus* (Hymenoptera: Aphelinidae) on *Bemisia tabaci* (Homoptera: Aleyrodidae). *Insect Sci.* 12: 461-465.
- Giron, D., Pincebourde, S., Casas, J., 2004. Lifetime gains of host-feeding in a synovigenic parasitic wasp. *Physiol. Entomol.* 29, 436-442.
- Greenberg, S.M., Jones, W.A., Liu, T.X., 2002. Interactions among two species of *Eretmocerus* (Hymenoptera: Aphelinidae), two species of whiteflies (Homoptera: Aleyrodidae), and tomato. *Environ. Entomol.* 31: 397-402.
- Greenberg, S.M., Jones, W.A., Liu, T.X., 2008. Bemisia tabaci (Homoptera: Aleyrodidae) instar effects on rate of parasitism by Eretmocerus mundus and Encarsia pergandiella (Hymenoptera: Aphelinidae). Entomol. Sci. 11: 97-103.
- Greenberg, S.M., Jones, W.A., Liu, T.X., 2009. Tritrophic interactions among host plants, whiteflies, and parasitoids. *Southwest. Entomol.* 34: 431-445.
- Gruenhagen, N.M., Perring, T.M., 2001a. Plant influences on silverleaf whitefly oviposition and development and the potential for enemy-free space. *Entomol. Exp. Appl.* 99: 387-391.
- Gruenhagen, N.M., Perring, T.M., 2001b. Impact of leaf trichomes on parasitoid behavior and parasitism of silverleaf whiteflies (Homoptera: Aleyrodidae). Southwest. Entomol. 26: 279-290.
- Gu, X.S., Bu, W.J., Xu, W.H., Bai, Y.C., Liu, B.M., Liu, T.X., 2008. Population suppression of *Bemisia tabaci* (Hemiptera: Aleyrodidae) using yellow sticky traps and *Eretmocerus* nr. *rajasthanicus* (Hymenoptera: Aphelinidae) on tomato plants in greenhouses. *Insect Sci.* 15: 263-270.
- Harvey, J.A., van Dam N.M., Gols, R., 2003. Interactions over four trophic levels: food plant quality affects development of a hyperparasitoid as mediated through a herbivore and its primary parasitoid. J. Anim. Ecol. 72: 520-531.
- Headrick, D.H., Bellows, Jr T.S., Perring, T.M., 1996a. Host-plant effects on the behavior of *Eretmocerus* sp. nr. *californicus* females raised on melon. *Entomophaga* 41: 15-26.
- Headrick, D.H., Bellows, Jr T.S., Perring, T.M., 1996b. Behaviours of female *Eretmocerus* sp. nr. *californicus* (Hymenoptera: Aphelinidae) attacking *Bemisia argentifolii* (Homoptera: Aleyrodidae) on cotton, *Gossypium hirsutum* (Malavaceae) and rockmelon, *Cucumis melo* (Cucurbitaceae). *Biol. Control* 6: 64-75.
- Heimpel, G.E., Collier, T.R. 1996. The evolution of host-feeding behavior in insect parasitoids. *Biol. Rev.* 71: 373-400.
- Heimpel, G.E., Rosenheim, J.A., 1995. Dynamic host feeding by the parasitoid *Aphytis melinus*: the balance between current and future reproduction. *J. Anim. Ecol.* 64: 153-167.
- Heinz, K.M., Nelson, J.M., 1996. Interspecific interactions among natural enemies of *Bemisia* in an inundative biological control program. *Biol.Control* 6: 384-393.
- Hu, X.S., Dennehy, T.J., Ni, X.Z., Zhao, H.Y., Nichols, R.L., Li, X.C., 2011. Potential adaptation of a Q biotype whitefly population from poinsettia to field crops. *Insect Sci.* 18: 719-728.
- Huang, Y., Loomans, A.J.M., van Lenteren, J.C., Xu, R.M., 2009. Hyperparasitism behavior of the autoparasitoid *Encarsia tricolor* on two secondary host species. *BioControl* 54: 411-424.
- Hunter, M.S., Collier, T.R., Kelly, S.E., 2002. Does an autoparasitoid disrupt host suppression provided by a primary parasitoid? *Ecology* 83: 1459-1469.
- Inbar, M., Gerling, D., 2008. Plant-mediated interactions between whiteflies, herbivores, and natural enemies. *Annu. Rev. Entomol.* 53: 431-448.
- Jackson, D.M., Farnham, M.W., Simmons, A.M., van Giessen, W.A., Elsey, K.D., 2000. Effects of planting pattern of collards on resistance to whiteflies (Homoptera: Aleyrodidae) and on parasitoid abundance. J. Econ. Entomol. 93: 1227-1236.
- Jervis, M.A., Kidd, N.A.C., 1986. Host-feeding strategies in hymenopteran parasitoids. Biol. Rev. 61: 395-434.
- Johnson, M.W., Toscano, N.C., Reynolds, H.T., Sylvester, E.S., Kido, K., Natwick, E.T., 1982. Whiteflies cause problems for southern California growers. *Calif. Agric.* 36: 24-26.
- Jones, W.A., Greenberg, S.M., 1999. Host instar suitability of *Bemisia argentifolii* (Homoptera: Aleyrodidae) for the parasitoid *Encarsia pergandiella* (Hymenoptera: Aphelinidae). J. Agric. Urban Entomol. 16: 49-57.

- Kidd, N.A.C., Jervis, M.A., 1989. The effects of host-feeding behavior on the dynamics of parasitoidhost interactions, and the implications for biological control. *Res. Popul. Ecol.* 31: 235-274.
- Lee, D.H., Nyrop, J.P., Sanderson, J.P., 2009. Attraction of *Trialeurodes vaporariorum* and *Bemisia argentifolii* to eggplant, and its potential as a trap crop for whitefly management on greenhouse poinsettia. *Entomol. Exp. App.* 133: 105-116.
- Lee, D.H., Nyrop, J.P., Sanderson, J.P., 2011. Avoidance of natural enemies by adult whiteflies, *Bemisia argentifolii*, and effects on host plant choice. *Biol. Control* 58: 302-309.
- Lenteren, J.C. van, Li, Z.H., Kamerman, J.W., Rumei, X., 1995. The parasite-host relationship between *Encarsia formosa* (Hymenoptera: Aphelinidae) and *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae) XXVI. Leaf hairs reduce the capacity of *Encarsia* to control greenhouse whitefly on cucumber. J. Appl. Entomol. 119: 553-559.
- Lenteren, J.C. van, Noldus, L.P.J.J., 1990. Whitefly-plant relationships: behavioral and ecological aspects of whitefly-plant relationships, in: Gerling D. (eds.), Whiteflies: Bionomics, Pest status and Management. Intercept, Andover, Hants, pp. 47-89.
- Li, S.J., Xue, X., Ahmed, M.Z., Ren, S.X., Du, Y.Z. et al., 2011. Host plants and natural enemies of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in China. *Insect Sci.* 18: 101-120.
- Lin, K., Wu, K.M., Zhang, Y.J., Guo, Y.Y., 2007. Overwintering and population dynamics of *Bemisia tabaci* biotype B in greenhouse during the spring in northern China. *Crop Prot.* 26: 1831-1838.
- Liu, S.S., Colvin, J., De Barro, P.J., 2012. Species concepts as applied to the whitefly *Bemisia tabaci* systematic: how many species are there? *J. Integr. Agric.* 11: 176-186.
- Liu, T.X., 2000. Population dynamics of *Bemisia argentifolii* (Homoptera: Aleyrodidae) of spring collard and relationship to yield in the Lower Rio Grande Valley of Texas. J. Econ. Entomol. 93: 750-756.
- Liu, T.X., 2003. Response of four melon varieties to silverleaf whitefly (Homoptera: Aleyrodidae) under laboratory and field conditions. Subtrop. *Plant Sci.* 55: 27-31.
- Liu, T.X., 2007. Life history of *Eretmocerus melanoscutus* (Hymenoptera: Aphelinidae) parasitizing nymphs of *Bemisia tabaci* biotype B (Homoptera: Aleyrodidae). *Biol. Control* 42: 77-85.
- Liu, T.X., Stansly, P.A., 1995. Oviposition by *Bemisia argentifolii* (Homoptera: Aleyrodidae): Effects of tomato leaf factors and insecticidal residues. J. Econ. Entomol. 88: 992-997.
- Lopez, S.N., Andornoa, A.V., 2009. Evaluation of the local population of *Eretmocerus mundus* (Hymenoptera: Aphelinidae) for biological control of *Bemisia tabaci* biotype B (Hemiptera: Aleyrodidae) in greenhouse peppers in Argentina. *Biol. Control* 50: 317-323.
- Luo, C., Liu, T.X., 2011. Fitness of *Encarsia sophia* (Hymenoptera: Aphelinidae) parasitizing *Trialeurodes vaporariorum* and *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Insect Sci.* 18: 84-91.
- Mallet, J., 1989. The evolution of insecticide resistance: Have the insect won? Tree 4: 336-340.
- Mansaray, A., Sundufu, A.J., 2010. Effect of three bean species on the development and reproduction of a population of the parasitoid, *Encarsia bimaculata*, on the whitefly, *Bemisia tabaci*. J. Insect Sci. 10: 28.
- Martin, J.H., Mifsud, D., Rapisarda, C., 2000. The whiteflies (Hemiptera: Aleyrodidae) of Europe and Mediterranean Basin. *Bull. Entomol. Res.* 90: 407-448.
- Mayer, R.T., Inbar, M., McKenzie, C.L., Shatters, R., Borowicz, V. et al., 2002. Multitrophic interactions of the silverleaf whitefly, host plants, competing herbivores, and phytopathogens. *Arch. Insect Biochem. Physiol.* 51: 151-169.
- McAuslane, H.J., Simmons, A.M., Jackson, D.M., 2000. Parasitism of *Bemisia argentifolii* on collard with reduced or normal leaf wax. *Fla. Entomol.* 83: 428-437.
- Mound, L.A., Halsey, S.H., 1978. Whiteflies of the World. British Museum of Natural History, Wiley, New York, NY p. 340.
- Naranjo, S.E., Cañas, L., Ellsworth, P.C., 2009. Mortality and population dynamics of *Bemisia tabaci* within a multi-crop system, in: Mason P.G., Gillespie D.R., Vincent C.D. (Eds.), Proceedings of the Third International Symposium on Biological Control of Arthropods. Christchurch, New Zealand, Publ. FHTET-2008-06: USDA Forest Service, pp: 202-207.
- Naranjo, S.E., Ellsworth, P.C., 2005. Mortality dynamics and population regulation in *Bemisia tabaci*. Entomol. Exp. Appl. 116: 93-108.
- Naranjo, S.E., Ellsworth, P.C., 2009. The contribution of conservation biological control to integrated pest control of *Bemisia tabaci* in cotton. *Biol. Control* 51: 458-470.

- Naranjo, S.E., Hagler, J.R., Ellsworth, P.C., 2003. Improved conservation of natural enemies with selective management systems for *Bemisia tabaci* (Homoptera: Aleyrodidae) in cotton. *Biocontrol Sci. Tech.* 13: 571-587.
- Njoroge, P., Shibairo, S., Githiri, S., & Mburu, M. (2014). Effects of Nitrogen Application on Growth and Yield of Snowpeas (Pisum sativum). Asian Journal Of Applied Science And Engineering, 3(1), 35-43. Retrieved from http://journals.abc.us.org/index.php/ajase/article/view/Njoroge
- Omondi, A.B., Obeng-Ofori, D., Kyerematen, R.A., Danquah, E.Y., 2005. Host preference and suitability of some selected crops for two biotypes of *Bemisia tabaci* in Ghana. *Entomol. Exp. Appl.* 115: 393-400.
- Oster, N., 1995. Factors influencing the parasitization efficiency of the parasitic wasp *Encarsia transvena*. M.Sc. Thesis, Tel Aviv University, pp.76.
- Pang, S.T., Wang, L., Hou, Y.H., Shi, Z.H., 2011. Interspecific interference competition between Encarsia formosa and Encarsia sophia (Hymenoptera: Aphelinidae) in parasitizing Bemisia tabaci (Hemiptera: Aleyrodidae) on five tomato varieties. Insect Sci. 18: 92-100.
- Pedata, P.A., Giorgini, M., Guerrieri, E., 2002. Interspecific host discrimination and within host competition between *Encarsia formosa* and *E. pergandiella* (Hymenoptera: Aphelinidae), two endoparasitoids of whiteflies (Hemiptera: Aleyrodidae). *Bull. Entomol. Res.* 92: 521-528.
- Perring, T.M., 2001. The Bemisia tabaci species complex. Crop Prot. 20: 725-737.
- Qiu, B.L., Ren, S.X., Lin, L., Wang, X.M., 2004. Effects of release density of *Eretmocerus* sp. (Hymenoptera: Aphelinidae) on the control effectiveness of *Bemisia tabaci* (Homoptera: Aleyrodidae). *Entomol. J. East China* 13: 27-30.
- Rajam, B., Peter, C., David, B.V., 1988. Influence of host plants on the parasitism of *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) by *Encarsia* sp. Curr. Sci. 57: 1246-1247.
- Rivero, A., Casas, J., 1999. Rate of nutrient allocation to egg production in a parasitic wasp. Proc. Biol. Sci. 266: 1169-1174.
- Royer, L., Fournet, S., Brunel, E., Boivin, G., 1999. Intra- and interspecific host discrimination by hostseeking larvae of coleopteran parasitoids. *Oecologia* 118: 59-68.
- Schuster, D.J., Mann, R.S., Toapanta, M., Cordero, R., Thompson, S. et al., 2010. Monitoring neonicotinoid resistance in biotype B of *Bemisia tabaci* in Florida. *Pest Manag. Sci.* 66: 186-195.
- Secker, A.E., Bedford, I.D., Markham, P.G., de Courcy-Williams, M.E., 1998. Squash, a reliable field indicator for the presence of the B biotype tobacco whitefly, *Bemisia tabaci*. British Crop Prot. Council, Brighton Conf. 3: 837-842.
- Shah, M.M.R., Liu, T.X., 2013. Feeding experience of *Bemisia tabaci* (Hemiptera: Aleyrodidae) affects their performance on different host plants. *PLoS ONE* 8, e77368.
- Shi, S.S., Zang, L.S., Liu, T.X., Yuan, C.C., Sun, G.Z., 2009. Host-feeding behavior of parasitoids on hosts and implication for biological control. *Acta Entomol. Sinica* 52: 424-433.
- Stansly, P.A., Calvo, J., Urbaneja, A., 2005a. Augmentative biological control of *Bemisia tabaci* biotype "Q" in Spanish greenhouse pepper production using *Eretmocerus* spp. *Crop Prot*. 24: 829-835.
- Stansly, P.A., Calvo, J., Urbaneja, A., 2005b. Release rates for control of *Bemisia tabaci* (Homoptera: Aleyrodidae) biotype "Q" with *Eretmocerus mundus* (Hymenoptera: Aphelinidae) in greenhouse tomato and pepper. *Biol. Control* 35: 124-133.
- Stansly, P.A., Sánchez, P.A., Rodríguez, J.M., Cañizares, F., Nieto, A. et al., 2004. Prospects for biological control of *Bemisia tabaci* (Homoptera, Aleyrodidae) in greenhouse tomatoes of southern Spain. Crop Prot. 23: 701-712.
- Ueno, T., 1999. Host-feeding and acceptance by a parasitic wasp (Hymenoptera: Ichneumonidae) as influenced by egg load and experience in a patch. *Evol. Ecol.* 13: 33-44.
- Urbaneja, A., Sánchez, E., Stansly, P.A., 2007. Life history of *Eretmocerus mundus*, a parasitoid of *Bemisia tabaci*, on tomato and sweet pepper. *BioControl* 52: 25-39.
- Videllet, P., Albajes, R., Gabarra, R., 1997. Host-feeding activity of *Encarsia pergandiella* Howard on *Bemisia tabaci* (Gennadius). *Bull. OILB/SROP.* 20: 147-152.
- Wan, F. H., Zhang, G. F., Liu, S. S., Luo, C., Chu, D. et al., 2009. Invasive mechanism and management strategy of *Bemisia tabaci* (Gennadius) biotype B: Progress Report of 973 Program on Invasive Alien Species in China. *Sci. China C Life Sci.* 52: 88-95.

- Wang, P., Sun, D.B., Qiu, B.L., Liu, S.S., 2011. The presence of six cryptic species of the whitefly *Bemisia tabaci* complex in China as revealed by crossing experiments. *Insect Sci.* 18: 67-77.
- Xiao, Y.F., Chen, J.J., Cantliffe, D., Mckenzie, C., Houben, K., Osborne, L.S., 2011. Establishment of papaya banker plant system for parasitoid, *Encarsia sophia* (Hymenoptera: Aphelinidae) against *Bemisia tabaci* (Hemiptera: Aleyrodidae) in greenhouse tomato production. *Biol. Control* 58: 239-247.
- Xiao, Y.F., Fadamiro, H.Y., 2009. Host preference and development of *Leptoglossus zonatus* (Hemiptera: Coreidae) on *Satsuma mandarin. J. Econ. Entomol.* 102: 1908-1914.
- Xu, J., Lin, K.K., Liu, S.S., 2011. Performance on different host plants of an alien and an indigenous Bemisia tabaci from China. J. Appl. Entomol. 135: 771-779.
- Yang, N.W., Wan, F.H., 2011. Host suitability of different instars of *Bemisia tabaci* biotype B for the parasitoid *Eretmocerus hayati*. *Biol. Control* 59: 313-317.
- Ying, J., Jian, H., Rui-Yan, M.A., Ju-Cai, H., 2003. Host plant preferences of *Bemisia tabaci* Gennadius. *Insect Sci.* 10: 109-114.
- Zang, L.S., Liu, T.X., 2008. Host-feeding of three parasitoid species on *Bemisia tabaci* biotype B and implications for whitefly biological control. *Entomol. Exp. Appl.* 127: 55-63.
- Zang, L.S., Liu, T.X., 2009. Food-deprived host-feeding parasitoids kill more pest insects. *Biocontrol Sci. Tech.* 19: 573-583.
- Zang, L.S., Liu, T.X., 2010. Effects of food deprivation on host feeding and parasitism of whitefly parasitoids. *Environ. Entomol.* 39: 912-918.
- Zang, L.S., Liu, T.X., Wan, F.H., 2011b. Reevaluation of the value of autoparasitoids in biological control. *PLoS ONE* 6, e20324
- Zang, L.S., Liu, T.X., Zhang, F., Shi, S.S., Wan, F.H., 2011a. Mating and host density affect host feeding and parasitism in two species of whitefly parasitoids. *Insect Sci.* 18: 78-83.
- Zhang, G.F., Lü, Z.C., Wan, F.H., 2007. Detection of *Bemisia tabaci* (Homoptera: Aleyrodidae) remains in predator guts using a sequence-characterized amplified region marker. *Entomol. Exp. Appl.* 123: 81-90.

-- 0 --

#### AJASE!!!

"Speedy publication service, Online archives, Paperless, web-based peer review system, Open access policy, Indexing in world known citation databases, Global circulation, Broad international readership and authorship, Online submission system, Minimum publication charge"

# Asian Journal of Applied Science and Engineering

(Multidisciplinary peer reviewed international journal)

ISSN: 2305-915X (print); 2307-9584 (Online) ICV 5.20; SJIF 2.607; UIF 2.0476

## **Open Access Philosophy**



Under **Open Access Philosophy**, AJASE will not charge for the access of its journals. This will ensure that a large percentage of students, scholars, researchers and practitioners will be able to benefit from the research published through ABC journals. Moreover, this process will also enable authors' papers to receive a higher ranking. A greater number of people being able to access and consequently refer to papers will mean a **higher citations** and **Impact Factor** for ABC journals. Following are advantages of Open Access Philosophy:

1. The full text of all issues of AJASE is freely available to anyone, online.

2. Your research work will be indexed and abstracted in the internationally reputed databases and search engines immediately after publication.

3. Open Access increases the number of downloads, page views, citations etc. increasing the rate of dissemination of your research work manifold.

4. It is inferred from past researches that the papers published under "**Open Access Philosophy**" are four times more likely to be cited than the papers published under "**Non-Open Access Philosophy**"



#### **Peer Review Policy**



Paperless, web-based peer review system, professional and helpful suggestions from reviewers. Articles in this journal have undergone a rigorous blind peer review system, based on initial editor screening and involving incountry and international refereeing, ensures that articles meet the highest standards of quality. Most ABC journals have ISSN with IMPACT FACTORS. It facilitates our scholars, researchers, scientists, professors, corporates, governmental research agencies, librarians etc., in a more positive way in their research proceedings.

#### Faster Turnaround Time

Many journals take many months, even years to publish research. By the time papers are published, often they become outdated. AJASE publishes papers in the shortest possible time, without compromising on quality. This will ensure that the latest research is published, allowing readers to gain maximum benefit. We provide feedback instantaneously and furnish details of the outcome within about 5 - 6 working days of submission of your research paper. This enables research scholars to use their time effectively on the actual research rather than on the follow ups.

#### Strong International network & Collaboration

We have exposure to wide range of industries across geographies and worldwide connect through international colleagues and thereby the recognition. We work in collaboration with extremely creditable companies, academic institutions, reputed publication units, government bodies and research firms. By publishing with us, you join ABC Global Research Community of 50,000 scientists / researchers.

For Details- go through the link: <u>www.ajase.weebly.com</u>



- Off Pantai Dalam, Kuala Lampur, Malaysia
- Road # 4, Shyamoli, Dhaka-1207, Bangladesh
- 3900 Woodhue Place, Alexandria, VA 22309, USA

#### www.abcreorg.weebly.com / www.abcjournals.net

Asian Business Consortium (ABC) is a multi-disciplinary research, training, publishing, digital library supporting and service house. Though founded in 2010 as the Business and Computing organization of Asia, it was reconstituted as the ABC in 2011. It has been working for creating and nurturing talents in USA, Malaysia and Bangladesh since its inception. The objectives of consortium are solely centered round the welfare and humane attitude of the founders who enthusiastically took up this noble cause and materialized it with a view to promote research and educational activities for the encouragement of scholars to develop their knowledge, to publish their analysis oriented scientific researches in international Journals, books, the task of organizing workshops, seminars, conferences, training, personality development programs and allied services.

In addition to research activities, ABC provides a good number of scholarships to the poor and meritorious students at various levels of education throughout the world. It plays an important role in the field of research by funding research projects and publishing the research papers. This consortium will unquestionably become the mouth-piece of the dark horses and unacknowledged scholar whose endowed and commendable contributions shall be provided an outlet keeping in mind the greater good of the larger society of the world.

ABC runs the following international referred journals for creating a platform to share the thoughts of professionals, scholars and academicians throughout the world.

#### **ABC Publications (ABC Journals)**

- Asian Accounting and Auditing Advancement (4A Journal)
- Asian Business Review (ABR)
- Asian Journal of Applied Sciences and Engineering (AJASE)
- Global Disclosure of Economics and Business (GDEB)
- ABC Journal of Advanced Research (ABC-JAR)
- International Journal of Reciprocal Symmetry and Theoretical Physics (IJRSTP)
- American Journal of Trade and Policy (AJTP)
- Asian Journal of Humanity, Art and Literature (AJHAL)
- Malaysian Journal of Medical and Biological Research (MJMBR)
- Asia Pacific Journal of Energy and Environment (APJEE)
- Engineering International (EI)
- ABC Research Alert (Online)

Each journal home page provides specific information for potential authors and subscribers. Open access policy, the quick review process, rich editorial boards and quality publications have already made ABC Journals unique. **ABC Journals** are published under the direct supervisions of renowned academicians of the world.

Collaboration in Conference: ABC considers high-quality conference papers for publication. Please contact us for detailed information.

Collaboration in Publishing: If you like to start writing a book, propose a new journal or advertise in ABC journals, please feel free to contact us.

