Mutant *KRAS* Codon 12 and 13 Alleles in Patients With Metastatic Colorectal Cancer: Assessment As Prognostic and Predictive Biomarkers of Response to Panitumumab

Marc Peeters, Jean-Yves Douillard, Eric Van Cutsem, Salvatore Siena, Kathy Zhang, Richard Williams, and Jeffrey Wiezorek

A B S T R A C T

Marc Peeters, Antwerp University
Hospital, Edegem; Eric Van Cutsem,
University Hospital Gasthuisberg,
Leuven, Belgium; Jean-Yves Douillard,
Institut de cancérologie de l'OuestRené Gauducheau, Nantes, France;
Salvatore Siena, Ospedale Niguarda Ca'
Granda, Milan, Italy; Kathy Zhang,
Amgen, South San Francisco; Richard
Williams and Jeffrey Wiezorek, Amgen,
Thousand Oaks, CA.

Published online ahead of print at www.jco.org on November 26, 2012.

Supported by Amgen, Thousand Oaks, CA.

Presented at the 2011 European Multidisciplinary Cancer Congress, Stockholm, Sweden, September 23-27, 2011; 9th Annual American Society of Clinical Oncology (ASCO) Gastrointestinal Cancers Symposium, San Francisco, CA, January 19-21, 2012; 48th Annual Meeting of ASCO, Chicago, IL, June 1-5, 2012; and the 14th Annual World Congress on Gastrointestinal Cancer, Barcelona, Spain, June 27-30, 2012.

Authors' disclosures of potential conflicts of interest and author contributions are found at the end of this article.

Corresponding author: Marc Peeters, MD, PhD, Department of Oncology, Antwerp University Hospital, Wilrijkstraat 10, B-2650 Edegem, Belgium; e-mail: Marc.Peeters@uza.be.

© 2012 by American Society of Clinical Oncology

0732-183X/13/3106-759/\$20.00 DOI: 10.1200/JCO.2012.45.1492

Purpose

Panitumumab, a fully human monoclonal antibody targeting the epidermal growth factor receptor (EGFR), has demonstrated significant improvements in progression-free survival (PFS) in patients with wild-type *KRAS* metastatic colorectal cancer (mCRC) in studies 20050203 (first line), 20050181 (second line), and 20020408 (monotherapy). Mutations in *KRAS* codons 12 and 13 are recognized biomarkers that predict lack of response to anti-EGFR antibody therapies. This retrospective analysis of three randomized phase III studies assessed the prognostic and predictive impact of individual mutant *KRAS* codon 12 and 13 alleles.

Patients and Methods

Patients were randomly assigned 1:1 to FOLFOX4 (infusional fluorouracil, leucovorin, and oxaliplatin) in study 20050203, FOLFIRI (fluorouracil, leucovorin, and irinotecan) in study 20050181, or best supportive care in study 20020408 with or without panitumumab 6.0 mg/kg once every 2 weeks. In all, 441 (20050203), 486 (20050181), and 126 (20020408) patients with mutant *KRAS* codon 12 or 13 alleles were included in the analysis.

Results

No mutant *KRAS* allele in patients treated on the control arm emerged as a consistent prognostic factor for PFS or overall survival (OS). In addition, no mutant *KRAS* allele was consistently identified as a predictive factor for PFS or OS in patients receiving panitumumab treatment. Significant interactions for individual mutant *KRAS* alleles were observed only in study 20050203 with G13D negatively and G12V positively associated with OS in the panitumumab-containing arm. Pooled analysis indicated that only G12A was associated with a negative predictive effect on OS.

Conclusion

In this retrospective analysis, results across three treatment regimens suggest that patients with mutant *KRAS* codon 12 or 13 mCRC tumors are unlikely to benefit from panitumumab therapy. Currently, panitumumab therapy should be limited to patients with wild-type *KRAS* mCRC.

J Clin Oncol 31:759-765. © 2012 by American Society of Clinical Oncology

INTRODUCTION

KRAS is a small G protein that acts as a transducer in the epidermal growth factor receptor (EGFR) pathway. Acquired *KRAS* codon 12 or 13 gain-of-function mutations lead to constitutive signaling through the EGFR pathway and to downstream activation of MAPK- and PI3K-dependent pathways. In approximately 40% of all metastatic colorectal cancer (mCRC) tumors, one of several heterozygous *KRAS* codon 12 or 13 mutations is detected. In individual patients, these point mutations are frequently detected in both primary and

metastatic lesions,^{4,5} consistent with the notion that *KRAS* mutations are acquired early during colorectal tumorigenesis.

Panitumumab is a fully human monoclonal antibody that targets the extracellular region of EGFR and effectively blocks ligand-dependent signaling downstream of the receptor. In the first-line 20050203 study, the second-line 20050181 study, and the monotherapy 20020408 study, panitumumab significantly improved progression-free survival (PFS) and response rate in patients with wild-type (wt) *KRAS* mCRC but not in their mutant *KRAS* mCRC counterparts. Collectively, mutant

KRAS codon 12 and 13 alleles are established biomarkers for lack of response to anti-EGFR monoclonal antibodies in patients with mCRC. ^{6,7,9-11}

However, recent retrospective analyses^{12,13} have suggested that patients whose tumors harbor a specific *KRAS* exon 2 mutation, a glycine (G; single-letter amino acid code) to aspartate (D) mutation at codon 13 (G13D), may derive clinical benefit from an anti-EGFR monoclonal antibody therapy in chemorefractory settings and in first-line combination therapy with irinotecan or oxaliplatin. In this analysis of 1,053 patients with mutant *KRAS* codon 12 or 13 alleles, we retrospectively examined the seven most common mutant *KRAS* codon 12 and 13 alleles for their prognostic and predictive impact on outcomes in patients with mCRC receiving panitumumab-containing therapy across three randomized phase III studies.

PATIENTS AND METHODS

Data Sets

Studies 20050203, 20050181, and 20020408 were open-label, multicenter, controlled phase III trials. ⁶⁻⁸ Patients in these trials were randomly assigned 1:1 to receive FOLFOX4 (infusional fluorouracil, leucovorin, and oxaliplatin) in study 20050203, FOLFIRI (fluorouracil, leucovorin, and irinotecan) in study 20050181, or best supportive care (BSC) in study 20020408 with or without panitumumab 6.0 mg/kg intravenously every 2 weeks. In studies 20020408 and 20050203, randomization was stratified by geographic region and Eastern Cooperative Oncology Group (ECOG) performance status. The primary end point for both studies was PFS. Key secondary end points included overall survival (OS), response rate, and safety. In study 20050181, randomization was stratified by prior oxaliplatin treatment, prior bevacizumab treatment, and ECOG performance status. The coprimary end points were PFS and OS. Key secondary end points included response rate and safety.

For all three phase III trials, key eligibility criteria included age ≥ 18 years, metastatic adenocarcinoma of the colon or rectum, measurable disease, ECOG performance status of 0 to 2, no prior anti-EGFR therapy, and paraffinembedded tumor tissue available for central biomarker analyses. The *KRAS* status of patients' tumors was neither required nor assayed at study entry but was performed after all patients had been enrolled.

KRAS testing was conducted by a blinded central laboratory and determined by using the TheraScreen K-RAS Mutation Kit (Qiagen, Manchester, United Kingdom) that detects the seven most common mutations in KRAS codons 12 and 13 (KRAS G12A, G12C, G12D, G12R, G12S, G12V, and G13D). Individual KRAS allele testing was performed without knowledge of

patient clinical outcomes. Descriptive statistics were provided for patient demographics and baseline characteristics in studies 20050203 and 20050181 but were not conducted in study 20020408 because of the relatively low number of patients with each mutant KRAS allele.

Statistical Analysis

The primary objective of this study was to examine the prognostic and predictive impact of the seven most common mutations in KRAS codons 12 and 13 on PFS, OS, and response rate in patients with mCRC who received panitumumab or control therapy. The analysis was conducted separately for each KRAS allele, for each study, and for all three studies combined. For prognostic analyses, comparisons were made between the outcomes of patients whose tumors harbored a specific KRAS mutation and the remaining patients whose tumors harbored any of the remaining six mutant KRAS alleles. Prognostic analyses were performed exclusively on patients who received control therapy (ie, non-panitumumab-containing). For analyses of the predictive impact of mutant KRAS alleles, relative treatment effects of panitumumab-containing and non-panitumumab-containing therapies were estimated among patients whose tumors harbored wt KRAS, any of the indicated mutant KRAS alleles (analyzed together as a group), or the specified individual mutant KRAS allele. Hazard ratios (HRs) and 95% CIs for PFS and OS were obtained by using the Cox proportional hazards model. A descriptive quantitative interaction test¹⁴ was conducted to assess the relative treatment effect on PFS and OS between the specific mutant KRAS codon 12 or 13 allele and the other KRAS mutations. No adjustments were made for multiple testing. HRs were stratified by study for the pooled analysis. All statistical evaluations were performed with SAS software, version 11 (SAS Institute, Cary, NC).

RESULTS

Patients

KRAS status was ascertained in mCRC tumors from 1,096 (93%) of 1,183 patients in study 20050203 (panitumumab plus FOLFOX4 ν FOLFOX4 alone), 1,083 (91%) of 1,186 patients in study 20050181 (panitumumab plus FOLFIRI ν FOLFIRI alone), and 427 (92%) of 463 patients in study 20020408 (panitumumab plus BSC ν BSC). This analysis of patients with mutant *KRAS* codon 12 or 13 mCRC included 441 (40%) of 1,096 patients in study 20050203, 486 (45%) of 1,083 patients in study 20050181, and 126 (30%) of 427 patients in study 20020408. The distribution of mutant *KRAS* codon 12 and 13 alleles was conserved across these three phase III studies and was equally balanced between the treatment and control arms (Table 1).

 Table 1. Distribution of Patients With Mutant KRAS Codon 12 and 13 mCRC Included in the Current Analysis From Studies 20050203, 20050181, and 20020408, Segregated by Treatment Arm

					, 0 (, ,						
	Study 20050203				Study 20050181				Study 20020408			
	Pmab + FOLFOX4 (n = 221)		FOLFOX4 (n = 220)		Pmab + FOLFIRI (n = 238)		FOLFIRI (n = 248)		Pmab + BSC (n = 56)		BSC (n = 70)	
KRAS Allele	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
G12D	68	31	57	26	77	32	70	28	21	38	22	31
G12V	57	26	64	29	62	26	79	32	10	18	18	26
G13D	46	21	52	24	39	16	45	18	9	16	11	16
G12C	16	7	19	9	26	11	19	8	6	11	6	9
G12A	21	10	13	6	17	7	17	7	6	11	5	7
G12S	13	6	14	6	12	5	13	5	4	7	6	9
G12R	0	0	1	< 1	5	2	4	2	0	0	2	3

Abbreviations: BSC, best supportive care; FOLFIRI, fluorouracil, leucovorin, and irinotecan; FOLFOX4, infusional fluorouracil, leucovorin, and oxaliplatin; mCRC, metastatic colorectal cancer; Pmab, panitumumab.

The results are consistent with published mCRC KRAS mutation analysis ¹⁵⁻¹⁸ and are comparable to the more than 9,000 primary colon and rectum adenocarcinoma cases in the public Catalogue of Somatic Mutations in Cancer (COSMIC) mutation database. ¹⁹ Together, KRAS G12D, G12V, and G13D comprised more than 70% of all mutant KRAS codon 12 and 13 alleles in each of the three studies. KRAS G12R was detected in less than 2% of mutant KRAS tumors and was not analyzed further.

Baseline demographic and clinical features were generally balanced in all mutant *KRAS* allele subgroups in studies 20050203 (Appendix Fig A1, online only) and 20050181 (Appendix Fig A2, online only) with the percentage of white patients, ECOG performance status, primary tumor site, incidence of liver plus other metastatic sites, prior surgery, and intensity of study therapy being similar across *KRAS* allelic subgroups and by treatment arm.

Prognostic Impact of KRAS Alleles

To evaluate the prognostic impact of KRAS codon 12 and 13 mutations, HRs with corresponding 95% CIs were plotted for the non–panitumumab-containing control arms of the first-line (20050203),

second-line (20050181), and monotherapy (20020408) studies (Fig 1). HRs for patients whose tumors harbored each of the individual mutant alleles were ordered by allele frequency and were compared with the other mutant *KRAS* codon 12 and 13 alleles combined.

The 95% CIs for the calculated HRs did not cross unity for mutant *KRAS* allele G12C (HR, 2.06; 95% CI, 1.16 to 3.65), which appeared as a negative prognostic factor for PFS but not for OS in study 20050203. None of the mutant *KRAS* alleles in study 20050181 were associated with a prognostic impact. In study 20020408, the 95% CIs for the calculated HRs did not cross unity for alleles *KRAS* G12C (HR, 2.47; 95% CI, 1.04 to 5.90) and *KRAS* G12A (HR, 5.30; 95% CI, 1.96 to 14.34), which both appeared as negative prognostic factors for OS but not for PFS. Taken together, no single mutant *KRAS* allele was a consistent negative or positive prognostic factor for both PFS and OS or across lines of mCRC therapy.

Predictive Impact of KRAS Alleles on Panitumumab Efficacy

The predictive effect of mutant *KRAS* codon 12 and 13 alleles on PFS and OS was also evaluated in all three phase III panitumumab studies (Fig 2). HRs with 95% CIs were plotted for patients whose

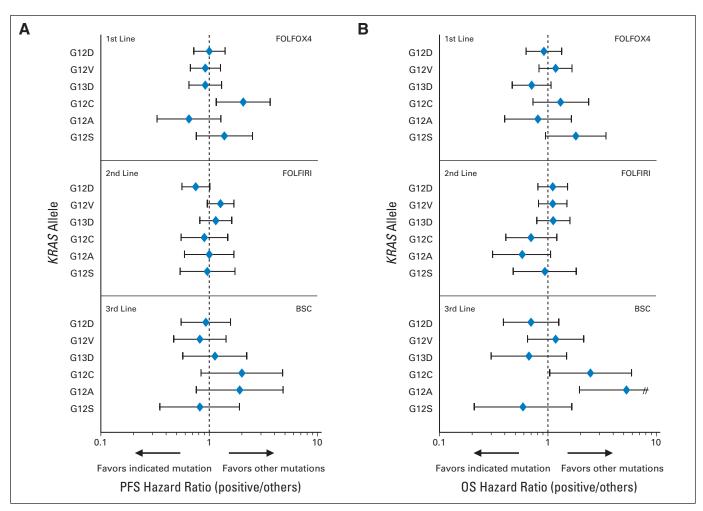


Fig 1. Prognostic impact of mutant KRAS codon 12 and 13 alleles (A) progression-free survival (PFS) and (B) overall survival (OS) in patients receiving control (non-panitumumab-containing) therapy. Point estimates for hazard ratios and their corresponding 95% Cls are plotted for the indicated mutant KRAS codon 12 and 13 alleles and are compared with the other mutant KRAS codon 12 and 13 alleles as a group. BSC, best supportive care; FOLFIRI, fluorouracil, leucovorin, and irinotecan; FOLFOX4, infusional fluorouracil, leucovorin, and oxaliplatin.

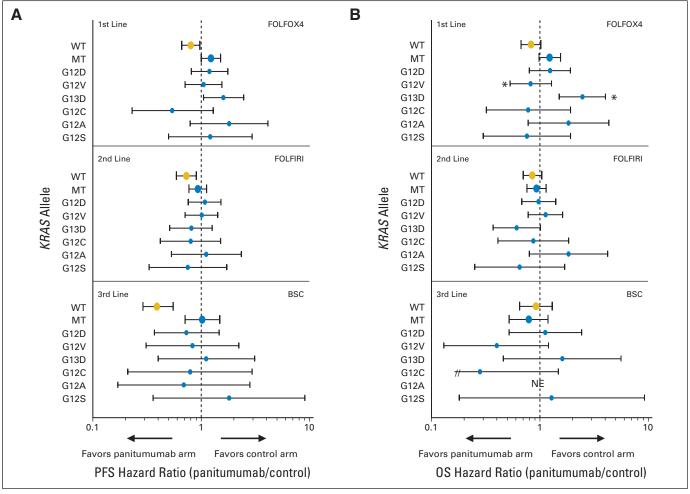


Fig 2. Predictive impact of mutant KRAS codon 12 and 13 alleles on (A) progression-free survival (PFS) and (B) overall survival (OS) in patients receiving either control (non-panitumumab-containing) or panitumumab-containing therapy. Point estimates for hazard ratios and their corresponding 95% CIs are plotted for wild-type (WT) KRAS and for the indicated mutant (MT) KRAS codon 12 and 13 alleles and are compared with the other mutant KRAS codon 12 and 13 alleles as a group. (*) Positive interaction test between indicated mutant KRAS allele and therapy. BSC, best supportive care; FOLFIRI, fluorouracil, leucovorin, and irinotecan; FOLFOX4, infusional fluorouracil, leucovorin, and oxaliplatin; NE, not estimable.

tumors harbored either wt KRAS or the indicated individual mutant KRAS alleles and were compared with the entire collection of mutant KRAS alleles. KRAS G13D was the only allele for which the 95% CIs for the calculated HRs did not cross unity, and it appeared as a negative predictive factor for both PFS (HR, 1.60; 95% CI, 1.05 to 2.46) and OS (HR, 2.47; 95% CI, 1.51 to 4.03) in the panitumumab-containing arm of the first-line 20050203 study. However, when a quantitative interaction test was conducted (Table 2), KRAS G13D was significantly associated only with a negative impact on OS (P = .0018) but not PFS (P = .1609). A borderline statistically significant positive impact on OS in study 20050203 was observed by interaction testing for KRAS G12V (P = .0369), but the 95% CIs for the calculated OS HR crossed unity (Fig 2). Taken together, across three studies, none of the individual mutant KRAS alleles were consistently associated with panitumumab treatment effects on PFS or OS outcomes. However, consistent with previous reports, ^{6,7,9} mutant KRAS alleles as a collective group were a negative predictive factor for both PFS and OS in panitumumab-containing therapies.

Response rates with 95% CIs were plotted for patients whose tumors harbored wt KRAS, any mutant KRAS codon 12 and 13 allele, or any indicated individual mutant KRAS allele (Appendix Fig A3, online only). In studies 20050203 and 20050181, objective responses for patients with mutant KRAS mCRC included only partial responses, and no complete responses were observed. 6,7 Response rates were similar across all mutant KRAS allele subgroups within each of the first- and second-line mCRC trials; 95% CIs for response rates of all individual mutant KRAS allele subgroups overlapped with each other, indicating no predictive effects of individual mutant KRAS codon 12 and 13 alleles on response rates. In study 20020408, no patient whose tumor harbored a mutant KRAS codon 12 or 13 allele responded to panitumumab therapy. As a collective group, mutant KRAS alleles were a negative predictive factor for response rate in the panitumumab-containing arms of all three trials.

Pooled Analysis of the Predictive Effect on PFS and OS by KRAS Alleles Across Panitumumab Studies

Pooled analysis of the three phase III trials was performed to increase sample size and to detect any significant trends that were not detectable in the individual studies. The analysis used individual patient-level data stratified by study. HRs and 95% CIs for PFS and OS

Table 2. P Values Determined From Quantitative Interaction Testing Exploring the Interaction Between the Specified Mutant KRAS Allele and Therapy on Either OS or PFS

KRAS	Study 20	050203	Study 20	0050181	Study 20020408		
Allele	OS	PFS	OS	PFS	OS	PFS	
G12D	.9870	.8692	.7351	.3658	.42	.41	
G12V	.0369*	.4229	.2449	.7023	.48	.56	
G13D	.0018*	.1609	.0665	.4736	.37	.90	
G12C	.3005	.0590	.8457	.6291	N/D†	N/D†	
G12A	.3362	.3279	.0974	.6547	N/D†	N/D†	
G12S	.2866	.9641	.4437	.5878	N/D†	N/D†	

Abbreviations: N/D, not determined; OS, overall survival; PFS, progression-free survival.

were plotted for 1,053 patients whose tumors harbored one of the *KRAS* codon 12 or 13 alleles pooled from studies 20050203, 20050181, and 20020408 (Fig 3). Only a single mutant *KRAS* allele, G12A, emerged as a predictive factor and was associated with a negative panitumumab treatment effect on OS but not on PFS. The earlier noted impacts of mutant *KRAS* G12V and *KRAS* G13D alleles on patient outcomes were no longer observed in the pooled analysis.

DISCUSSION

Preclinical studies have suggested that individual *KRAS* codon 12 or 13 alleles have displayed quantitative and/or qualitative differences in transforming capacity and other biologic phenotypes. Specifically, *KRAS* codon 12 mutations have displayed greater in vitro transforming ability when compared with *KRAS* codon 13 mutations, ²⁰⁻²² and individual mutant *KRAS* codon 12 alleles have had a differential impact on cellular transformation. ²³ Furthermore, the signaling networks activated downstream of individual mutant *KRAS* alleles have varied significantly. ^{22,23} Despite these intrinsic biologic differences observed in defined experimental systems, the differential prognostic

or predictive impact of individual mutant *KRAS* codon 12 or 13 alleles in a genetically complex and heterogeneous disease such as mCRC have remained untested by a systematic approach.

This study is the largest retrospective analysis evaluating the seven most common mutations in *KRAS* codons 12 and 13 for prognostic and predictive impact in patients with mCRC receiving an anti-EGFR therapy. Enrollment was completed in trials 20050203, 20050181, and 20020408 before *KRAS* was established as a predictive marker for outcomes in patients with mCRC. *KRAS* allele status was ascertained in more than 90% of the patients in each of the three phase III trials. A total of 1,053 patients were included in this analysis from these three open-label, multicenter, randomized, controlled phase III trials. The frequency and distribution of mutant *KRAS* codon 12 and 13 alleles were conserved across the trials, equally balanced between the treatment and control arms, and consistent with public domain data and prior publications. ¹⁵⁻¹⁹ Baseline demographics and clinical features were also balanced by treatment arm and comparable between all mutant *KRAS* allelic subgroups in trials 20050203 and 20050181.

Analysis of mutant *KRAS* codon 12 and 13 alleles on prognosis in the control arms of the three phase III trials suggested a trend toward a negative prognostic factor for *KRAS* G12C on PFS for patients receiving FOLFOX4 and on OS for patients receiving BSC. A trend as a negative prognostic factor was also observed for *KRAS* G12A on OS for patients receiving BSC. However, no single mutant *KRAS* allele was a consistent prognostic factor on both PFS and OS or across lines of mCRC therapy.

The prognostic significance of *KRAS* mutations has been assessed in a multitude of studies, with conflicting results.²⁴ Several studies have suggested that *KRAS* mutations are a negative prognostic indicator in CRC.²⁵ When considering the five largest studies, a prognostic impact was reported by four of these studies.²⁶ The RASCAL II metanalysis indicated that *KRAS* G12V in Duke's C CRC patients was associated with a significant reduction in disease-free survival and OS.^{17,27} Samowitz et al²⁸ performed the first population-based study on *KRAS* mutations in CRC, and results suggested that mutations in *KRAS* codon 13 were associated with poor OS. In addition, De Roock et al¹² recently reported that *KRAS* G13D mCRC tumors had worse OS compared with wt *KRAS* tumors and compared with tumors

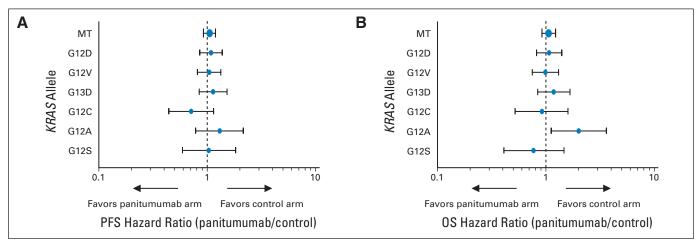


Fig 3. Pooled analysis of studies 20050203, 20050181, and 20020408: Predictive impact of mutant (MT) KRAS codon 12 and 13 alleles on (A) progression-free survival (PFS) and (B) overall survival (OS) in patients receiving either control (non–panitumumab-containing) or panitumumab-containing therapy. Point estimates for hazard ratios and their corresponding 95% Cls are plotted for the indicated mutant KRAS codon 12 and 13 alleles and are compared with the other mutant KRAS codon 12 and 13 alleles as a group.

^{*}Quantitative interaction tests with P < .05.

[†]Not performed because of limiting number of patients in these KRAS allele subgroups.

bearing other KRAS mutations, although significance was lost in multivariate analysis. 12

In contrast, a large number of reported studies (reviewed in Klump et al²⁶) have found no association of *KRAS* gene mutations with survival, either in isolation or in combination with other tumor suppressor genes. Interpretation of the various studies published on the prognostic role of *KRAS* in mCRC may be challenging because of the different *KRAS* mutations investigated and variations in data collection, staging techniques, and mutant *KRAS* detection methodologies. In patients with mCRC treated with an anti-EGFR monoclonal therapy, it has been suggested that *KRAS* mutations are unlikely to be prognostic (independent of any specific treatment) and are likely predictive (attributable to treatment).²⁹

In the analysis reported here, there was no consistent evidence that any individual mutant *KRAS* allele, compared with the remaining mutant *KRAS* alleles or the entire mutant *KRAS* group, had a differential impact on response rate, PFS, or OS. Only in the first-line FOLFOX4 treatment setting of study 20050203 were statistically significant differences observed for individual mutant *KRAS* alleles: *KRAS* G12V was favorably and *KRAS* G13D was unfavorably associated with panitumumab treatment effects on OS but not on PFS or response rate. Because associations with OS were observed only in the FOLFOX4 treatment setting and because other *KRAS* mutations have been associated with platinum sensitivity, ³⁰ it is possible that selected mutant *KRAS* alleles may have a differential impact on patient outcomes in the specific context of coadministration with oxaliplatincontaining chemotherapy.

High *KRAS* ascertainment rates and consistent *KRAS* testing methodology permitted the pooling of data from all three phase III trials to potentially identify predictive trends across three lines of therapy that may not have been observed from analysis of any single trial. Pooled analyses indicated that no individual mutant *KRAS* codon 12 and 13 allele was associated with outcomes for both PFS and OS, relative to other *KRAS* mutations. A trend was observed for *KRAS* G12A, which was associated with a negative panitumumab predictive effect only on OS. *KRAS* alleles G12V and G13D were no longer associated with outcomes in the pooled analysis, suggesting there were no predictive trends across lines of therapy.

These results are in contrast with reported cetuximab data, 12,13 which have suggested patients with *KRAS* G13D responded to an anti-EGFR monoclonal antibody therapy. However, this improved survival in patients with *KRAS* G13D was not significant in the cetuximab monotherapy arm, and therefore the confounding effect of the chemotherapy backbone cannot be excluded. 12 In addition, a recent retrospective analysis of 110 patients treated with cetuximab 31 reported that patients whose tumor harbored a *KRAS* G13D allele did not benefit from cetuximab treatment (n = 12) and had a trend toward lower OS compared with patients whose tumors harbored either wt *KRAS* or one of the other *KRAS* mutations. Although pani-

tumumab and cetuximab recognize similar epitopes,³² they are of different antibody isotypes and may have different abilities to bind to EGFR mutations.³³ It is unclear whether these or other characteristics may have contributed to the conflicting results reported between these antibodies.

The cetuximab studies 12,13,31 and the analysis reported here were limited by their retrospective nature, they used subset analysis, and were subject to chance observations. None of the studies made adjustments for multiple testing. Other possible limitations were the low frequency and low number of patients in the specific mutant *KRAS* allelic subgroups, such as in the De Roock et al study which had a total of 32 patients in the *KRAS* G13D subgroup in pooled analyses of patients in cetuximab monotherapy (n = 10) and cetuximab plus chemotherapy studies (n = 22).

On the basis of all of the available data and consistent with current clinical treatment guidelines, we suggest that patients with mCRC tumors that harbor any of the most common mutant *KRAS* codon 12 or 13 alleles are unlikely to benefit from panitumumab therapy. Therefore, only mCRC patients with wt *KRAS* tumors should be treated with panitumumab therapy.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Although all authors completed the disclosure declaration, the following author(s) and/or an author's immediate family member(s) indicated a financial or other interest that is relevant to the subject matter under consideration in this article. Certain relationships marked with a "U" are those for which no compensation was received; those relationships marked with a "C" were compensated. For a detailed description of the disclosure categories, or for more information about ASCO's conflict of interest policy, please refer to the Author Disclosure Declaration and the Disclosures of Potential Conflicts of Interest section in Information for Contributors. Employment or Leadership Position: Kathy Zhang, Amgen (C); Richard Williams, Amgen (C); Jeffrey Wiezorek, Amgen (C) Consultant or Advisory Role: Marc Peeters, Amgen (C); Jean-Yves Douillard, Amgen (C), Merck Serono (C); Salvatore Siena, Amgen (C), Roche (C), sanofi-aventis (C) Stock Ownership: Kathy Zhang, Amgen; Richard Williams, Amgen; Jeffrey Wiezorek, Amgen Honoraria: Marc Peeters, Amgen; Jean-Yves Douillard, Amgen, Merck Serono Research Funding: Marc Peeters, Amgen; Jean-Yves Douillard, Merck Serono; Eric Van Cutsem, Amgen Expert Testimony: None Other Remuneration: Marc Peeters, Amgen

AUTHOR CONTRIBUTIONS

Conception and design: Salvatore Siena, Kathy Zhang, Jeffrey Wiezorek Provision of study materials or patients: Eric Van Cutsem Collection and assembly of data: Kathy Zhang, Richard Williams Data analysis and interpretation: All authors Manuscript writing: All authors Final approval of manuscript: All authors

REFERENCES

- 1. Bardelli A, Siena S: Molecular mechanisms of resistance to cetuximab and panitumumab in colorectal cancer. J Clin Oncol 28:1254-1261, 2010
- 2. Siena S, Sartore-Bianchi A, Di Nicolantonio F, et al: Biomarkers predicting clinical outcome of epidermal growth factor receptor-targeted therapy

in metastatic colorectal cancer. J Natl Cancer Inst 101:1308-1324, 2009

- **3.** Roth AD, Tejpar S, Delorenzi M, et al: Prognostic role of KRAS and BRAF in stage II and III resected colon cancer: Results of the translational study on the PETACC-3, EORTC 40993, SAKK 60-00 trial. J Clin Oncol 28:466-474, 2010
- **4.** Knijn N, Mekenkamp LJ, Klomp M, et al: KRAS mutation analysis: A comparison between primary tu-

mours and matched liver metastases in 305 colorectal cancer patients. Br J Cancer 104:1020-1026, 2011

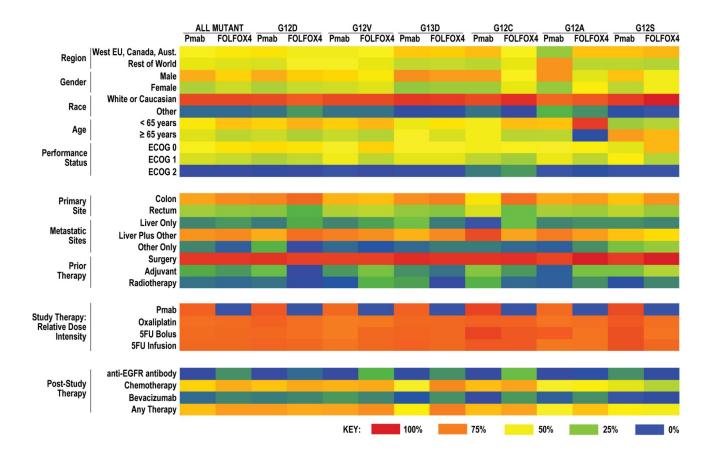
- **5.** Santini D, Loupakis F, Vincenzi B, et al: High concordance of KRAS status between primary colorectal tumors and related metastatic sites: Implications for clinical practice. Oncologist 13:1270-1275, 2008
- **6.** Douillard JY, Siena S, Cassidy J, et al: Randomized, phase III trial of panitumumab with infusional fluorouracil, leucovorin, and oxaliplatin

- (FOLFOX4) versus FOLFOX4 alone as first-line treatment in patients with previously untreated metastatic colorectal cancer: The PRIME study. J Clin Oncol 28:4697-4705, 2010
- 7. Peeters M, Price TJ, Cervantes A, et al: Randomized phase III study of panitumumab with fluorouracil, leucovorin, and irinotecan (FOLFIRI) compared with FOLFIRI alone as second-line treatment in patients with metastatic colorectal cancer. J Clin Oncol 28:4706-4713, 2010
- 8. Van Cutsem E, Peeters M, Siena S, et al: Open-label phase III trial of panitumumab plus best supportive care compared with best supportive care alone in patients with chemotherapy-refractory metastatic colorectal cancer. J Clin Oncol 25:1658-1664, 2007
- 9. Amado RG, Wolf M, Peeters M, et al: Wildtype KRAS is required for panitumumab efficacy in patients with metastatic colorectal cancer. J Clin Oncol 26:1626-1634, 2008
- 10. Khambata-Ford S, Garrett CR, Meropol NJ, et al: Expression of epiregulin and amphiregulin and K-ras mutation status predict disease control in metastatic colorectal cancer patients treated with cetuximab. J Clin Oncol 25:3230-3237, 2007
- 11. Van Cutsem E, Köhne CH, Hitre E, et al: Cetuximab and chemotherapy as initial treatment for metastatic colorectal cancer. N Engl J Med 360:1408-1417, 2009
- 12. De Roock W, Jonker DJ, Di Nicolantonio F, et al: Association of KRAS p.G13D mutation with outcome in patients with chemotherapy-refractory metastatic colorectal cancer treated with cetuximab. JAMA 304:1812-1820, 2010
- 13. Tejpar S, Celik I, Schlichting M, et al: Association of KRAS G13D tumor mutations with outcome in patients with metastatic colorectal cancer treated with first-line chemotherapy with or without cetuximab. J Clin Oncol 30:3570-3577, 2012
- 14. Gail M. Simon R: Testing for qualitative interactions between treatment effects and patient subsets. Biometrics 41:361-372, 1985

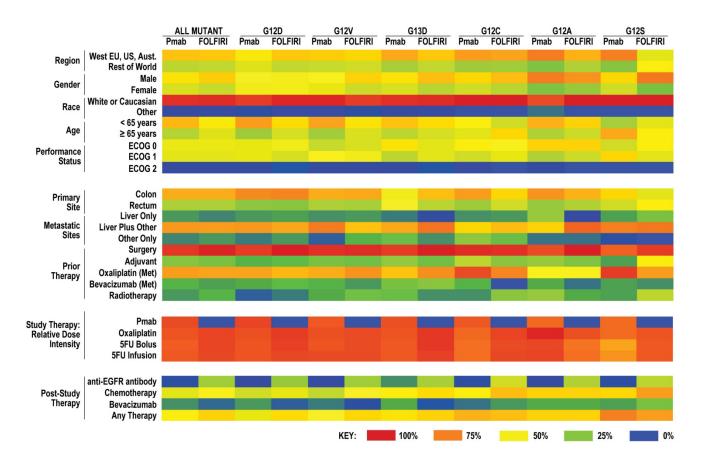
- 15. Deschoolmeester V, Boeckx C, Baay M, et al: KRAS mutation detection and prognostic potential in sporadic colorectal cancer using high-resolution melting analysis. Br J Cancer 103:1627-1636, 2010
- 16. Neumann J, Zeindl-Eberhart E, Kirchner T, et al: Frequency and type of KRAS mutations in routine diagnostic analysis of metastatic colorectal cancer. Pathol Res Pract 205:858-862 2009
- 17. Andreyev HJ, Norman AR, Cunningham D, et al: Kirsten ras mutations in patients with colorectal cancer: The 'RASCAL II' study. Br J Cancer 85:692-696 2001
- 18. Sundström M, Edlund K, Lindell M, et al: KRAS analysis in colorectal carcinoma: Analytical aspects of pyrosequencing and allele-specific PCR in clinical practice. BMC Cancer 10:660, 2010
- 19. Forbes SA, Bindal N, Bamford S, et al: COSMIC: Mining complete cancer genomes in the Catalogue of Somatic Mutations in Cancer, Nucleic Acids Res. 39:D945-D950, 2011
- 20. Guerrero S, Casanova I, Farré L, et al: K-ras codon 12 mutation induces higher level of resistance to apoptosis and predisposition to anchorageindependent growth than codon 13 mutation or proto-oncogene overexpression. Cancer Res 60: 6750-6756 2000
- 21. Guerrero S, Figueras A, Casanova I, et al: Codon 12 and codon 13 mutations at the K-ras gene induce different soft tissue sarcoma types in nude mice, FASEB J 16:1642-1644, 2002
- 22. Smith G, Bounds R, Wolf H, et al: Activating K-Ras mutations outwith 'hotspot' codons in sporadic colorectal tumours: Implications for personalised cancer medicine. Br J Cancer 102:693-703,
- 23. Céspedes MV, Sancho FJ, Guerrero S, et al: K-ras Asp12 mutant neither interacts with Raf, nor signals through Erk and is less tumorigenic than K-ras Val12. Carcinogenesis 27:2190-2200, 2006
- 24. Deschoolmeester V, Baay M, Specenier P, et al: A review of the most promising biomarkers in

- colorectal cancer: One step closer to targeted therapy. Oncologist 15:699-731, 2010
- 25. Locker GY, Hamilton S, Harris J, et al: ASCO 2006 update of recommendations for the use of tumor markers in gastrointestinal cancer. J Clin Oncol 24:5313-5327, 2006
- 26. Klump B, Nehls O, Okech T, et al: Molecular lesions in colorectal cancer: Impact on prognosis? Original data and review of the literature. Int J Colorectal Dis 19:23-42, 2004
- 27. Russo A, Bazan V, Agnese V, et al: Prognostic and predictive factors in colorectal cancer: Kirsten Ras in CRC (RASCAL) and TP53CRC collaborative studies. Ann Oncol 16:iv44-iv49, 2005
- 28. Samowitz WS, Curtin K, Schaffer D, et al: Relationship of Ki-ras mutations in colon cancers to tumor location, stage, and survival: A populationbased study. Cancer Epidemiol Biomarkers Prev 9:1193-1197, 2000
- 29. De Roock W, De Vriendt V, Normanno N, et al: KRAS, BRAF, PIK3CA, and PTEN mutations: Implications for targeted therapies in metastatic colorectal cancer, Lancet Oncol 12:594-603, 2011
- 30. Orlandi A, Di Salvatore M, Basso M, et al: ERCC1, KRAS mutation, and oxaliplatin sensitivity in colorectal cancer: Old dogs and new tricks. J Clin Oncol 30, 2012 (suppl 4; abstr 489)
- 31. Gajate P, Sastre J, Bando I, et al: Influence of KRAS p.G13D mutation in patients with metastatic colorectal cancer treated with cetuximab. Clin Colorectal Cancer 11:291-296, 2012
- 32. Freeman D, Sun J, Bass R, et al: Panitumumab and cetuximab epitope mapping and in vitro activity. J Clin Oncol 15s:630s, 2008 (suppl; abstr 14536)
- 33. Montagut C, Dalmases A, Bellosillo B, et al: Identification of a mutation in the extracellular domain of the epidermal growth factor receptor conferring cetuximab resistance in colorectal cancer. Nat Med 18:221-223, 2012

Supplementary Fig 1.



Supplementary Fig 2.



Supplementary Fig 3.

